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Remarks:

This application was filed on 17 - 06 - 1998 as a divisional application to the application mentioned under INID code 62.

A process for the purification of phosphatidylserine (54)

(57) The invention relates to a process for the purification of phosphatidylserines (PS) by selective extraction of PS-containing phospholipid mixtures in diphasic systems of organic solvents and, eventually, by further crystallization.

Description

[0001] The present invention relates to a process for the purification of phosphatidylserines of formula (I), hereinafter referred to as PS, by relying on the different partition coefficients in diphasic organic solvents.

[0002] The compounds which may be subjected to the process of the invention have the general formula (I) and are prepared starting preferentially from compounds (II), according to the procedure of European Patent Application n. 95119395.9-51212:

wherein R¹ and R², which are the same or different, are selected from a saturated, mono- or polyunsaturated C_{10} - C_{80} acyl group;

X = OH or OM wherein M is an alkali, alkaline-earth metal, ammonium or alkylammonium;

R₃ = CH₂CH₂NH₂ or CH₂CH₂ N⁺(CH₃)₃.

[0003] The importance of the compounds (I) is various, particularly in the preparation of pharmaceutical compositions for the therapy of involutive cerebral syndromes of different origin, such as vascular pathologies on atheroschlerotic base or not and/or serille decline; for the preparation of liposomial formulations and more recently for dietetic compositions comprising natural lecithins, particularly soy lecithin enriched in phosphatidyl-L-serine, hereinafter referred to as PS(L), containing mostly polyunsaturated tathy acids as alory residues.

[0004] The increasing demand for industrial amounts of PS(L) at a reasonable cost prompted the Applicant to carry out a thorough investigation to fulfill such a need.

[0005] Comfurius P et al.. Biochim. Biophys. Acta 488, 36 (1977) first discloses the production of an about 1/1 mixture of PS(L) and phosphatidic acid (PA). by reacting under pressure at 45°C and at pH 5.6, in a diphasic ethyl ether/water system, egg lecitini or synthetic phosphatidylcholines with L-serine in the presence of partially purified PLD enzyme (from achbase).

40 [0006] PS(L) is then purified by chromatography on cellulose using a chloroform/methanol mixture as eluent. It is evident that this procedure is not suited to an industrial production both due to the use of ethyl ether and the low selectivity; furthemore the chromatographic procedure disclosed in this purification method is further ground against the industrial applicability of said methods for the use of chloroform in the eluent mixture, the low productivity and high cost.

[0007] The invention provides a method for the purification of the PS, relying on the different partition coefficients in 45 diphasic organic solvent systems of phosphatides such as PS, PA, phosphatidychloline (PC), phosphatidylethanolamine (PE) and the corresponding lysophosphatides in the form of the corresponding salts, particularly the corresponding calcium salts in the diphasic heptane/methanol system.

[0008] It is possible to increase, in a 90% yield, the purify of a PS(L) obtained from Epikuron 200^(M) from 88% to 95% thanks to its preferential ripartition in the heptane phase; similarly, the purify of a PS(L) obtained from Epikuron 135^(M) was increased from 55% to about 80%. Finally, a further purification of PS can be obtained by crystallization from heptane/acetione in the form of the calcium salt and subsequent conversion into any other salt, according to conventional stephologies.

[0009] The process of the invention can be conveniently applied for the purification of phosphatidyl-(L)-serines wherein R₁ and R₂ are acyl chairs of palmitic, stearic, oleic, linoleic acids in similar proportions to that of say leathin or wherein R₁ and R₂ are acyl chairs of palmitic, stearic, palmitolic, oleic, linoleic, arenchlooric acids in similar proportions to that of egg lecithin or wherein R₁ and R₂ are the same acyl chains in similar proportions to that of starting material

[0010] The procedures reported in the following further exemplify the invention.

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Example 1

Preparation of PS(L) starting from soy lecithin Epikuron 200

5 [0011] 20 g d Epituron 200^(R) (Lucas Meyre) and 100 ml of toluene are placed into a 1.000 ml reactor, under introgen, and the solution is concentrated under vacuum distilling about 80 ml of the solvent. Fresh toluene is added and the solution is concentrated again under reduced pressure. The procedure is repeated until reaching a content in ethanol or other C₁-C₂ alcobios, which are usually present in commercial lectifin, below 20 ppm. The residue is taken up into fresh toluene to a volume of 400 ml and added with 9.5 g of (L) eleme. The resulting suspension is added with the aqueous solution (300 ml) containing PLD rom ATCC 55717, prepared according to the procedures of European Patient Application N 9119390.5-1212 (Example 1) and having an enzymatic activity of 2 Urin, added at 10°C with 3.34 g of acid under which 21°C below 10°C with 3.45 g of a cide under the case of the procedure of Everyment of the case of the c

Example 2

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Preparation of PS(L) starting from soy lecithin Epikuron 135

28 (0012) 400 Kg of Epikuron 135⁽⁶⁾ (Lucas Meyer), 3000 I of boluene, 100 I of water are placed into a 5,000 I stainless steel reactor, under intigogan, and the mixture is concentrated under vacuum distilling at 45°C about 1,000 I of solvants. Another 6,000 I stainless steel reactor is loaded with 1,355 I of fermentation broth from ATCC 55717, containing about 3, AIVI of PLD, 22.7 kg of calcium chloride, 27.6 kg of sodium acetate Inthipyrate, and at I/O C2 I of 80% acetic acid 625 kg of L-serine (final pl4 4.2). The two solutions are combined and the resulting mixture is heated to and kept at 25°C 30 with strong stirring for 5h. HPLC analysis shows a PS(L) content of about 75% of the total phospholipids. The mixture is then added with a suspension of 35 kg of decastile in 500 I of toluene and filtered, washing the tilter with 400 I of toluenewater (37), V/V). The acueous phase is then separated and treated to recover (L)-serine whereas the organic phase, after further filtration on decalite, is concentrated under vacuum to an about 440 kg residue, which is taked, with 500 toluened and the state of the contractive of the contractive and the contractive of the contracti

[0013] The product is further purified by treatment with 2,000 I of acetone and dried, to give about 273 kg of PS(L) calcium salt (58%).

[0014] A 20 g sample was purified by extraction with heptane/methanol, analogously to what described in example 1, to give 11,6 g of PS(L) calcium salt (80%).

Example 3

Preparation of PS(L) starting from egg lecithin.

46 [0015] 13 g of Ovothin 160^(R) (60% PC; Lucas Meyer), 158 ml of toluene and 38 g of (L)-serine are placed into a 500 ml reactor, under nitrogen. The resulting suspension is added with the aqueous solution (300 ml) contain IP pLD from ATCC 55717, and having an enzymatic activity of 2 U/ml, acted at 10°C with 1.4 g of calcium chloride, 1.7 g of sodium accelate trihydrate and glacial acetic acid necessary to obtain a pH of about 4.1. The resulting diphrasic system is heated to a temperature of 25° ±2°C and kept under strong stiming to about 6.1. The muture is then filtered on decalite into the strong straing for about 6.1. The muture is the nitred on decalite into its first the washed with 2x 100 ml of 10 louene; the organic phase is separated from the acqueous phase, containing the serine excess, and concentrated under reduced pressure to give a residue which is taken up into 320 ml of in heptand 100 ml of methand. The lower methanol phase is discarded whereas the higher one is diluted with 35 ml of heptane and further extracted with 95 ml of methanol. The higher phase is separated, concentrated under vacuum to small organic une and added under stirring at 5°C with 250 ml of acetone to give, upon filtration and drying under vacuum, 7.3 g of 5° PS(L) calcium salt with a 45% FHLC.

Example 4

Preparation of DLPS(L) starting from DLPC.

5 [0016] Repeating the procedure described in example 1, but using 20 g of L-a-dilinoneylphosphatidylcholine, referred to as DLPC, instead of 20 g of Epikuron 200⁽⁶⁾, 15.1 g of L-a-dilinoneylphosphatidyl-L-serine, referred to as DLPS(L), as the calcium salt (96% HPLC purity), are obtained.

Claims

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1. A process for the purification of the compounds of formula (I)

wherein:

 R^1 and R^2 , which are the same or different, are selected from saturated, mono- or polyunsaturated C_{10} - C_{20} acyl group;

X = OH or OM, wherein M is an alkali, alkaline-earth metal, ammonium or alkylammonium; by selective extraction of PS-containing phospholipid mixtures in diphasic systems of organic solvents.

- A process according to claim 1, wherein phosphatidy/serine is purified from the other phospholipids, particularly PC, PE, PA and the corresponding lysophospholipids, by selective extraction of the corresponding calcium salt from an heptane/methanol mixture.
- A process for the further purification of phosphatidylserine obtained according to claim 2, wherein the phosphatidylserine is subjected to a crystallization from heptane/acetone, in the form of the calcium salt, and subsequent conversion into any other salt, according to conventional techniques.

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EUROPEAN SEARCH REPORT

Application Number EP 98 11 1111

	DOCUMENTS CONSIDER				
ategory	Citation of document with indica of relevant passage	tion, where appropriate,	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.6)	
A	US 5 084 215 A (KEARNS 28 January 1992 * claims 1,3 *	JOHN J ET AL)	1-3	C07F9/10	
A	COMFURIUS P ET AL: "I SYNTHESIS OF PHOSPHATI PURIFICATION BY CM-CEI CHROMATOGRAPHY" BIOCHIMICA ET BIOPHYSI vol. 488, no. 1, 20 Ju 36-42, XP000603420 * page 40, paragraph 2	DYLSERINE AND LULOSE COLUMN CA ACTA, lly 1977, pages	1-3		
				TECHNICAL FIELDS SEARCHED (Int.Cl.6)	
				SEARCHED (Int.CI.6)	
	The present search report has been	drawn up for all claims			
Place of search		Date of completion of the search	· '	Examiner	
THE HAGUE		22 April 1999	Bes	Beslier, L	
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ANNEX TO THE EUROPEAN SEARCH REPORT ON EUROPEAN PATENT APPLICATION NO.

EP 98 11 1111

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in oway faible for these particulars which are merely given for the purpose of information.

22-04-1999

	Patent document cited in search report		Publication date	Patent family member(s)		Publication date	
U	S 5084215	A	28-01-1992	US US	4814111 4714571	A A	21-03-1989 22-12-1989
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For more details about this annex -see Official Journal of the European Patent Office, No. 12/82